

# Package ‘ModCon’

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**Type** Package

**Title** Modifying splice site usage by changing the mRNP code, while maintaining the genetic code

**Version** 1.12.0

**Description** Collection of functions to calculate a nucleotide sequence surrounding for splice donors sites to either activate or repress donor usage. The proposed alternative nucleotide sequence encodes the same amino acid and could be applied e.g. in reporter systems to silence or activate cryptic splice donor sites.

**License** GPL-3 + file LICENSE

**Encoding** UTF-8

**LazyData** true

**VignetteBuilder** knitr

**Depends** data.table, parallel, utils, stats, R (>= 4.1)

**Suggests** testthat, knitr, rmarkdown, dplyr, shinycssloaders, shiny, shinyFiles, shinydashboard, shinyjs

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calculateHZEIint	<i>Calculate HZEI integral of nucleotide sequence</i>
------------------	---

---

### Description

This function calculates the HZEI integral of a nucleotide sequence.

### Usage

```
calculateHZEIint(ntSequence)
```

### Arguments

ntSequence	Character value of nucleotide sequence whose HZEI integral will be calculated. It should be at least 11 nt long and only contain bases 'A', 'G', 'C', 'T'.
------------	--

### Value

Integer value stating the HZEI integral of the given sequence ntSequence

## Examples

```
## Example to increase HZEI integral for a given coding sequence
x <- calculateHZEIint('ATACCAGCCAGCTATTACATTT')
```

---

calculateMaxEntScanScore

*Calculate MaxEntScan score of a splice site sequence*

---

## Description

This function calculates the MaxEntScan score of either splice donor (SD) or acceptor sequences (SA).

## Usage

```
calculateMaxEntScanScore(seqVector, ssType)
```

## Arguments

seqVector	Character value of nucleotide sequence of a splice site sequence. SA sequences should be 23nt long (20 intronic, 3 exonic) and SD sequences should be 9nt long (3 exonic, 6 intronic). Only bases 'A', 'G', 'C', 'T' permitted.
ssType	Numeric value which indicates the type of splice site. Either '3' for an SA or '5' for an SD.

## Value

Numeric vector stating the MaxEntScan score per splice site sequence entered with seqVector

## Examples

```
calculateMaxEntScanScore('TTCCAACGAACTTTTGTAGGGA', 3)
calculateMaxEntScanScore('GAGGTAAGT', 5)
```

---

cds                                      *CDS of firefly luciferase*

---

**Description**

Character string of the nucleotide sequence encoding the firefly luciferase.

**Usage**

cds

**Format**

character string

**Examples**

cds

---

changeSequenceHZEI            *Adjust HZEI integral of nucleotide sequence*

---

**Description**

Adjust the HZEI integral of a nucleotide sequence (min. 24nt long)

**Usage**

```
changeSequenceHZEI(inSeq, increaseHZEI=TRUE, nGenerations=50, parentSize=300,
startParentSize=1000, bestRate=50, semiLuckyRate=20, luckyRate=5, mutationRate=1e-04,
optiRate=100, sdMaximalHBS=10, acMaximalMaxent=4, nCores=-1)
```

**Arguments**

inSeq	Character value of nucleotide sequence (min 24nt long, only bases A, G, T or C)
increaseHZEI	Logical value if HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.
nGenerations	Numeric value setting maximal number of generations
parentSize	Numeric value setting size of parent generations, generated from previous generations
startParentSize	Numeric value setting size of initiated parent generation of sequences
bestRate	Numeric value setting percentage how many of the fittest sequences are used to produce the next generation

semiLuckyRate	Numeric value setting percentage of sequences which are selected for breeding with a probability based on the respective HZEI-score integral
luckyRate	Numeric value setting percentage of sequences which are randomly selected for breeding
mutationRate	Numeric value setting chance of each codon, to mutate randomly within a child sequence
optiRate	Numeric value setting level of HZEI integral optimization
sdMaximalHBS	Numeric value of minimal HBS of SDs which should be tried to be degraded in their intrinsic strength
acMaximalMaxent	Numeric value of minimal MaxEntScan score of SAs which should be tried to be degraded in their intrinsic strength
nCores	Numeric value setting number of cores which should be used for parallel computations. If set to '-1' all available cores are selected.

**Value**

Character value of a nucleotide sequence encoding the same amino acid sequence as inSeq, but an increased HZEI integral, due to alternative codon selection.

**Examples**

```
## Load R packages
library('parallel')
library('utils')
library('data.table')

## Set parameters for genetic algorithm
inSeq <- 'ATGGAAGACGCCAAAACATAAAGAAAGGCCCGCCATTCTATCCGCTGGAAGATGGAACC'

## Increase HZEI integral
res <- changeSequenceHZEI(inSeq)

## Setting additional parameters
res <- changeSequenceHZEI(inSeq, increaseHZEI=TRUE, nGenerations=50, parentSize=300,
startParentSize=1000, bestRate=50, semiLuckyRate=20, luckyRate=5, mutationRate=1e-04,
optiRate=100, sdMaximalHBS=10, acMaximalMaxent=4, nCores=1)

## Access sequence with highest generated HZEI intregal
res[[3]]
```

---

Codons

*Table of codons and encoded amino acids*

---

**Description**

Table of codons and encoded amino acids

**Usage**

Codons

**Format**

A data frame with columns:

**ndiff** Indicator, how many codons encode the same amino acid

**AA** Amino acid three-lette code

**name** Amino acid full name

**seq** Codon sequence

**Examples**

Codons

---

createCodonMatrix      *Create codon matrix from coding nucleotide sequence*

---

**Description**

This function creates a codon matrix with 2 rows and as many columns as codons within the sequence.

**Usage**

```
createCodonMatrix(cds)
```

**Arguments**

**cds**                      Character value of nucleotide sequence whose HZEI integral will be calculated. It should be at least 3 nt long and only contain bases 'A', 'G', 'C', 'T'. Length must be a multiple of 3.

**Value**

Character matrix holding the encoded codon sequence in both rows.

**Examples**

```
## Example to create codon matrix  
createCodonMatrix("ATGAATGATCAAAAAGCTAGCC")
```

---

`createFilialSequencePopulation`*Generate new sequences by recombination*

---

**Description**

This function generates new sequences from set of parental sequences through recombination.

**Usage**

```
createFilialSequencePopulation(sequenceVector, generateNrecombinedSequences)
```

**Arguments**

`sequenceVector` Character vector of nucleotide sequences which will be used to create new sequences through recombination.

`generateNrecombinedSequences`

Numeric value setting number of recombined sequences which will be generated

**Value**

Character vector of nucleotide sequences, generated by recombination from the entered `sequenceVector`, holding as much filial sequences as stated in `generateNrecombinedSequences`. Modes of recombination are cross-over, insertion and random.

**Examples**

```
createFilialSequencePopulation(c('AAABBBCCDDDEEEFFF', 'GGGHHIIJJJKKLLL'), 3)
```

---

`decreaseGTsiteStrength`*Remove or degrade intrinsic strength of specific GT site while keeping the HZEI integral neutral.*

---

**Description**

Degrade or remove specific GT site from a coding sequence by codon selection keeping the HZEI integral near zero.

**Usage**

```
decreaseGTsiteStrength(cds, sdSeqStartPosition)
```

**Arguments**

`cds` Character value of a coding nucleotide sequence which holds the splice site of interest. Sequence length must be dividable by 3 and only contain bases 'A', 'G', 'C', 'T'.

`sdSeqStartPosition` Numeric value of position of the first nucleotide of the splice donor of interest

**Value**

Character vector of a nucleotide sequence encoding the same amino acid as the entered `cds`, but the intrinsic strength of a specific GT site within the CDS is degraded as much as possible.

**Examples**

```
library(data.table)
cds <- paste0('ATGGAAGACGCCAAAAACATAAAGAAAGCCCGGCCATTCTATCCGCTGGAAGATGGAACCGCTGGAGAGCAACTGCA',
' TAAGGCTATGAAGAGATACGCCCTGGTTCCTGGAACAATTGCTTTTACAGATGCACATATCGAGGTGGACATCACTTACGCTGAGTACTTCGAAA',
' TGCCGTTTCGGTTGGCAGAAGCTATGAAACGATATGGGCTGAATACAAATCACAGAATCGTCGTATGCAGTGAAAACCTCTTCAATTCTTTAT',
' GCCGGTGTGGGCGCCTTATTTATCGGAGTTGCAGTTGCGCCCGCAACGACATTATAATGAACGTGAATTGCTCAACAGTATGGGCATTTTCG',
' CAGCTACCGTGGTGTTCGTTTCCAAAAGGGGTTGCAAAAAATTTTGAACGTGCAAAAAAGCTCCCAATCATCCAAAAATATTATCATGG',
' ATTCTAAAACGGATTACCAGGGATTTTCAGTCGATGTACACGTTTCGTACATCTCATCTACCTCCCGTTTTAATGAATACGATTTTGTGCCAGA',
' GTCCTTCGATAGGACAAGACAATTGCACTGATCATGAACCTCTGGATCTACTGGTCTGCCTAAAGGTGTCGCTCGCCTCATAGAAGTCC',
' TGCCTGAGATTCTCGCATGCCAGAGATCCTATTTTTGGCAATCAAATCATTCCGGATACTGCGATTTAAGTGTTGTTCCATTCCATCACGGT',
' TTGGAATGTTTACTACACTCGGATATTTGATATGTGGATTTTCGAGTCGTCTTAATGTATAGATTTGAAGAAGAGCTGTTTCTGAGGAGCCTCA',
' GGATTACAAGATTCAAAGTGCCTGCTGGTGCCAACCTATTCTCTTCTTCGCCAAAAGCACTCTGATTGACAAATACGATTTATCTAATTTA',
' CACGAAATTGCTTCTGGTGGCGCTCCCCTCTCTAAGGAAGTCGGGGAAGCGGTTGCCAAGAGGTTCCATCTGCCAGGTATCAGGCAAGGATATG',
' GGCTCACTGAGACTACATCAGCTATTCTGATTACACCGAGGGGGATGATAAACCGGGCGCGTCCGGTAAAGTTGTTCCATTTTTTGAAGCGAA',
' GGTGTGGATCTGGATACCGGAAAACGCTGGGCGTTAATCAAAGAGGCGAACTGTGTGTGAGAGGTCCATGATTATGTCCGGTTATGTAAC',
' AATCCGGAAGCGACCAACGCCTTGATTGACAAGGATGGATGGCTACATTCTGGAGACATAGCTTACTGGGACGAAGACGAACACTTCTTCATCG',
' TTGACCGCTGAAGTCTCTGATTAAGTACAAAGGCTATCAGGTGGCTCCCGCTGAATTGGAATCCATCTTGTCCAACACCCCAACATCTTCGA',
' CGCAGGTGTCGAGGTCTTCCCGACGATGACGCCGGTGAACCTCCCGCCCGGTTGTTGTTTGGAGCACGGAAGAGCATGACGGAAAAAGAG',
' ATCGTGGATTACGTCGCCAGTCAAGTAAACCCGCAAAAAGTTGCGCGGAGGAGTTGTGTTTGGAGCAAGTACCGAAAGGTTTACCAGGAA',
' AACTCGACGCAAGAAAAATCAGAGAGATCCTCATAAAGGCCAAGAAGGGCGGAAAGATCGCCGTG')

sdSeqStartPosition <- 1001
cdsNew <- decreaseGTsiteStrength(cds, sdSeqStartPosition)
print(cdsNew)
```

---

degradeSAs

*Remove or degrade intrinsic strength of splice acceptors while adjusting HZEI integral.*

---

**Description**

Degrade or remove splice acceptor sites of certain intrinsic strength (in MaxEntScan score) from a coding sequence by codon selection while keeping the HZEI integral up.



**Usage**

```
degradeSAs(fanFunc, maxhbs=10, maxME=4, increaseHZEI=TRUE)
```

**Arguments**

fanFunc	codon matrix with two rows (see example below)
maxhbs	Numeric treshold which strength of internal donor sites should be degraded (in HBS)
maxME	Numeric treshold which strength of internal acceptor sites should be degraded (in MaxEntScan score)
increaseHZEI	Logical value if HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.

**Value**

Character value of a nucleotide sequence encoding the same amino acid as the entered codon matrix fan, but the intrinsic strength of all present splice acceptor (SA) sites is degraded as much as possible, in case they exceed the given treshold maxME. Additionally, splice donor site strengths greater maxhbs are avoided, during SA degradation.

**Examples**

```
library(data.table)
sdMaximalHBS <- 10
acMaximalMaxent <- 4
increaseHZEI <- TRUE
## Initiaing the Codons matrix plus corresponding amino acids
ntSequence <- 'TTTTGTCTTTTCTGTGTGGCAGTGGGATTAGCCTCCTATCGATCTATGCGATA'
## Create Codon Matrix by splitting up the sequence by 3nt
fanFunc <- createCodonMatrix(ntSequence)
degradeSAs(fanFunc, maxhbs=sdMaximalHBS, maxME=acMaximalMaxent, increaseHZEI=increaseHZEI)
```

---

degradeSDs	<i>Remove or degrade intrinsic strength of splice donors while adjusting HZEI integral.</i>
------------	---

---

**Description**

Degrade or remove splice donor sites of certain intrinsic strength (in HBS) from a coding sequence by codon selection.

**Usage**

```
degradeSDs(fanFunc, maxhbs=10, increaseHZEI=TRUE)
```

**Arguments**

fanFunc	Codon matrix with two rows (see example below)
maxhbs	Numeric treshold which strength of internal donor sites should be degraded
increaseHZEI	Logical value of HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.

**Value**

Character value of a nucleotide sequence encoding the same amino acid as the entered codon matrix fanFunc, but the intrinsic strength of all present splice donors (SD) sites is degraded as much as possible, in case they exceed the given treshold maxhbs.

**Examples**

```
library(data.table)
## Initiating the Codons matrix plus corresponding amino acids
ntSequence <- 'TTTTCGATCGGGATTAGCCTCCAGGTAAGTATCTATCGATCTATGCGATAG'
## Create Codon Matrix by splitting up the sequence by 3nt
fanFunc <- createCodonMatrix(ntSequence)
degradeSDs(fanFunc, maxhbs=10, increaseHZEI=TRUE)
```

---

```
generateRandomCodonsPerAA
```

*Randomly choose Codon to encode amino acid sequence*

---

**Description**

Encode amino acid sequence by random codon selection

**Usage**

```
generateRandomCodonsPerAA(aaVector)
```

**Arguments**

aaVector	Character vector of amino acids in three letter code (e.g. Met)
----------	---

**Value**

Character value of a nucleotide sequence encoding the same amino acid as the entered by aaVector by random Codon selection.

**Examples**

```
generateRandomCodonsPerAA(c('Lys', 'Lys'))
```

---

`getOverlappingVectorsFromVector`  
*Create overlapping subvectors*

---

**Description**

Create overlapping subvectors from large vector

**Usage**

`getOverlappingVectorsFromVector(largeVector, subvectorLength, subvectorOverlap )`

**Arguments**

`largeVector`      Large character vector to break down into overlapping subvectors  
`subvectorLength`      Numeric value of length of smaller subvectors  
`subvectorOverlap`      Numeric value of length of subvector overlap

**Value**

Creates a list of overlapping subvectors from an input vector `largeVector`. The length of these overlapping subvectors is stated by `subvectorLength` and the overlap of the resulting subvectors is stated by `subvectorOverlap`.

**Examples**

`getOverlappingVectorsFromVector(c(1,2,3,4), 2, 1)`

---

`hbg`      *Donor sequences and their HBS*

---

**Description**

Donor sequences and their HBS

**Usage**

`hbg`

**Format**

A data frame with columns:

**seq** 11nt long donor sequence

**hbs** HBS of the donor sequence

**special\_seq** Shorter version of the donor sequence

**Examples**

hbg

---

hex

*Hexamers and Z scores*

---

**Description**

Hexamers and Z scores

**Usage**

hex

**Format**

A data frame with columns:

**seq** Sequence of the hexamer.

**value** ZEI-score of the hexamer from HEXplorer.

**first** First codon within the hexamer.

**second** Second codon within the hexamer.

**first\_AA** First encoded amino acid within the hexamer (three letter code).

**second\_AA** Second encoded amino acid within the hexamer (three letter code).

**AA** Both encoded amino acid within the hexamer

**Examples**

hex

---

increaseGTsiteStrength

*Increasing intrinsic strength of specific GT site while keeping the HZEI integral neutral.*

---

## Description

Increasing intrinsic strength specific GT site from a coding sequence by codon selection keeping the HZEI integral near zero.

## Usage

```
increaseGTsiteStrength(cds, sdSeqStartPosition)
```

## Arguments

cds                    Coding nucleotide sequence which holds the splice site of interest  
sdSeqStartPosition        Numeric value of position of the first nucleotide of the splice donor of interest

## Value

Character vector of a nucleotide sequence encoding the same amino acid as the entered cds, but the intrinsic strength of a specific GT site within the CDS is enhanced as much as possible.

## Examples

```
library(data.table)
cds <- paste0('ATGGAAGACGCCAAAAACATAAAGAAAGGCCCGGCCATTCTATCCGCTGGAAGATGGAACCGCTGGAGAGCAACTGCA',
              'TAAGGCTATGAAGAGATACGCCCTGGTTCTGGAACAATTGCTTTTACAGATGCACATATCGAGGTGGACATCACTTACGCTGAGTACTTCGAAA',
              'TGCCGTTTCGGTTGGCAGAAGCTATGAAACGATATGGGCTGAATACAAATCACAGAATCGTCGATGCAGTGAAAACTCTCTCAATTCTTTAT',
              'GCCGGTGTGGCGCGTTATTTATCGGAGTTGCAGTTGCGCCCCGGAACGACATTTATAATGAACGTGAATTGCTCAACAGTATGGGCATTTTCG',
              'CAGCCTACCGTGGTGTTCGTTTCCAAAAAGGGTTGCAAAAAATTTTGAACGTGCAAAAAAGCTCCCAATCATCCAAAAAATTATTATCATGG',
              'ATTCTAAAACGGATTACCAGGGATTTTCAGTCGATGTACACGTTTCGTCACATCTCATCTACCTCCCGTTTTAATGAATACGATTTTGTGCCAGA',
              'GTCCTTCGATAGGGACAAGACAATTGCACTGATCATGAACTCCTCTGGATCTACTGGTCTGCCTAAAGGTGTCGCTCTGCCTCATAGAACTGCC',
              'TGCGTGAGATTCTCGCATGCCAGAGATCCTATTTTGGCAATCAAATCATTCCGGATACTGCGATTTAAGTGTGTGCCATTCCATCACGGTT',
              'TTGGAATGTTTACTACTCGGATATTTGATATGTGGATTTCGAGTCGCTTAATGTATAGATTTGAAGAAGAGCTGTTTCTGAGGAGCCTTCA',
              'GGATTACAAGATTCAAAGTGCCTGCTGGTGCCAACCCTATTCCTTCTTCGCCAAAAGCACTCTGATTGACAAATACGATTTATCTAATTTA',
              'CACGAAATGCTTCTGGTGGCGCTCCCTCTCTAAGGAAGTCGGGGAAGCGGTTGCCAAGAGGTTCCATCTGCCAGGTATCAGGCAAGGATATG',
              'GGCTCACTGAGACTACATCAGCTATTCTGATTACACCCGAGGGGGATGATAAACGGGCGCGGTCGGTAAAGTTGTTCCATTTTTGAAGCGAA',
              'GGTTGTGGATCTGGATACCGGAAAAACGCTGGGCGTTAATCAAAGAGGCGAACTGTGTGTGAGAGGTCCTATGATTATGTCCGGTTATGTAAC',
              'AATCCGGAAGCGACCAACGCCTTGATTGACAAGGATGGATGGCTACATTCTGGAGACATAGCTTACTGGGACGAAGACGAACACTTCTTCATCG',
              'TTGACCGCCTGAAGTCTCTGATTAAGTACAAAGGCTATCAGGTGGCTCCCGCTGAATTGGAATCCATCTTGCTCCAACACCCCAACATCTTCGA',
              'CGCAGGTGTCGAGGTCCTCCCGACGATGACGCCGGTGAACCTCCCGCCGCGTTGTTGTTTTGGAGCACGAAAAGACGATGACGGAAAAAGAG',
              'ATCGTGATTACGTCGCCAGTCAAGTAACAACCGGAAAAAGTTGCGCGGAGGAGTTGTGTTTTGGACGAAGTACCGAAAGGCTTACCGGAA',
              'AACTCGACGCAAGAAAAATCAGAGAGATCCTCATAAAGGCCAAGAAGGGCGGAAAGATCGCCGTG')

sdSeqStartPosition <- 1001
```

```

cdsNew <- increaseGTsiteStrength(cds, sdSeqStartPosition)
print(cdsNew)

```

---

ModCon

*ModCon*


---

### Description

Execute ModCon on a donor site within a coding sequence either increasing or decreasing its HZEI weight.

### Usage

```

ModCon(cds, sdSeqStartPosition, upChangeCodonsIn=16, downChangeCodonsIn=16,
optimizeContext=TRUE, sdMaximalHBS=10, acMaximalMaxent=4, optiRate=100,
nGenerations=50, parentSize=300, startParentSize=1000, bestRate=40,
semiLuckyRate=20, luckyRate=5, mutationRate=1e-04, nCores=-1)

```

### Arguments

cds	Character value of coding nucleotide sequence which holds the splice site of interest
sdSeqStartPosition	Numeric value of the position of the first nucleotide of the splice donor of interest
upChangeCodonsIn	Numeric value of number of codons to change upstream of the donor site of interest
downChangeCodonsIn	Numeric value of number of codons to change downstream of the donor site of interest
optimizeContext	Character value which determines, if TRUE (the default) the donor context will be adjusted to increase the splice site HEXplorer weight (SSHW), if FALSE, the SSHW will be decreased.
sdMaximalHBS	Numeric value of minimal HBS of SDs which should be tried to be degraded in their intrinsic strength
acMaximalMaxent	Numeric value of minimal MaxEntScan score of SAs which should be tried to be degraded in their intrinsic strength
optiRate	Numeric value setting level of HZEI integral optimization
nGenerations	Numeric value setting maximal number of generations
parentSize	Numeric value setting size of parent generations, generated from previous generations
startParentSize	Numeric value setting size of initiated parent generation of sequences

bestRate	Numeric value setting percentage how many of the fittest sequences are used to produce the next generation
semiLuckyRate	Numeric value setting percentage of sequences which are selected for breeding with a probability based on the respective HZEI-score integral
luckyRate	Numeric value setting percentage of sequences which are randomly selected for breeding
mutationRate	Numeric value setting chance of each codon, to mutate randomly within a child sequence
nCores	Numeric value setting number of cores which should be used for parallel computations. If set to '-1' all available cores are selected.

### Value

Creates a character value of a coding nucleotide sequence encoding the same amino acid sequence as the entered cds, but with an alternative nucleotide surrounding around the splice donor (SD) sequence position, stated with sdSeqStartPosition. Depending on the entered optimizeContext, the SD surrounding is either adjusted aiming to enhance or decrease the splice site HEXplorer wheigth.

### Examples

```
## Load R packages
library('parallel')
library('utils')
library('data.table')

## Set parameters for simplest use of ModCon (optimizing to 100%)
cds <- paste0('ATGGAAGACGCCAAAAACATAAAGAAAGGCCCGGCCATTCTATCCGCTGGAAGATGGAACCGCTGGAGAGCAACTGCA',

'TAAGGCTATGAAGAGATACGCCCTGGTTCCTGGAACAATTGCTTTTACAGATGCACATATCGAGGTGGACATCACTTACGCTGAGTACTTCGAAA',
'TGTCGGTTCGGTTGGCAGAAGCTATGAAACGATATGGGCTGAATACAAATCACAGAATCGTCGATGTCAGTGAAAACCTCTTCAATTCTTTAT',
'GCCGGTGTGGGCGCGTTATTTATCGGAGTTGCAGTTGCGCCCCGCAACGACATTTATAATGAACGTGAATTGCTCAACAGTATGGGCATTTCCG',
'CAGCCTACCGTGGTGTTCGTTTCCAAAAAGGGTTGCAAAAAATTTTGAACGTGCAAAAAAGCTCCCAATCATCCAAAAATTTATTCATGG',
'ATTCTAAAACGGATTACCAGGGATTTTCAGTCGATGTACAGTTCGTCACATCTCATCTACCTCCCGGTTTTAATGAATACGATTTTGTGCCAGA',
'GTCCCTCGATAGGGACAAGCAATTGCACTGATCATGAACTCCTCTGGATCTACTGGTCTGCCATAAAGGTGTCGCTCTGCCTCATAGAACTGCC',
'TGCGTGAGATTCTCGCATGCCAGAGATCCTATTTTTGGCAATCAAATCATTCCGGATACTGCGATTTAAGTGTTGTTCCATTCCATCACGGTT',
'TTGGAAATGTTTACTACACTCGGATATTTGATATGTGGATTTGAGTCGTCTTAATGTATAGATTTGAAGAAGAGCTGTTTCTGAGGAGCCTTCA',
'GGATTACAAGATTCAAAGTGCCTGCTGGTGCCAACCCTATTCTCTTCTCGCCAAAAGCACTCTGATTGACAAATACGATTTATCTAATTTA',
'CACGAAATGCTTCTGGTGGCGCTCCCCTCTTAAGGAAGTCGGGGAAGCGGTTGCCAAGAGGTTCCATCTGCCAGGTATCAGGCAAGGATATG',
'GGCTCACTGAGACTACATCAGCTATTCTGATTACACCCGAGGGGATGATAAACCGGGCGCGTCCGTAAGGTTGTTCCATTTTTGAAGCGAA',
'GGTGTGGATCTGGATACCGGAAAACGCTGGGCGTTAATCAAAGAGGCGAACTGTGTGTGAGAGGTCCTATGATTATGTCGGTTATGTAAC',
'AATCCGGAAGCGCAACGCCTTGATTGACAAGGATGGATGGCTACATTCTGGAGACATAGCTTACTGGGACGAAGACGAACACTTCTTCATCG',
'TTGACCGCCTGAAGTCTCTGATTAAGTACAAAGGCTATCAGGTGGCTCCCGCTGAATTGGAATCCATCTTGTCCAACACCCCAACATCTTCGA',
'CGCAGGTGTCGAGGTTCTCCGACGATGACGCCGGTGAACCTCCCGCCCGGTTGTTGTTTTGGAGCACGGAAGACGATGACGGAAAAAGAG',
'ATCGTGGATTACGTCGCCAGTCAAGTAACAACCGCAAAAAGTTGCGCGGAGGAGTTGTGTTTGTGGACGAAGTACCGAAAGGTTTACCGGAA',
'AACTCGACGCAAGAAAAATCAGAGAGATCCTCATAAAGGCAAGAAGGGCGGAAAGATCGCCGTG')

## Execute ModCon
finalSequence <- ModCon(cds, 1001)
```

```

## Print final cds sequence with the alternative SD nucleotide surrounding
print(finalSequence)

## More parameters can be set for use of ModCon when not optimizing to 100% (e.g. 50%)

## Execute ModCon
finalSequence <- ModCon(cds, 1001, upChangeCodonsIn=16, downChangeCodonsIn=16,
optimizeContext=FALSE, sdMaximalHBS=10, acMaximalMaxent=4,
optiRate=50, nGenerations=5, parentSize=200, startParentSize=800,
bestRate=50, semiLuckyRate=10, luckyRate=5, mutationRate=1e-03, nCores=1)

## Print final cds sequence with the alternative SD nucleotide surrounding
print(finalSequence)

```

---

mutatePopulation	<i>Randomly exchange codons within a set of sequences.</i>
------------------	--

---

## Description

For every codon within a set of nucleotide sequences randomly exchange the codon encoding the same amino acid to a certain chance.

## Usage

```
mutatePopulation(sequenceVector, codonReplacementChance)
```

## Arguments

sequenceVector Character vector of nucleotide sequences (at least 3 nt long)

codonReplacementChance

Numeric value of chance of a codons within the sequences to get exchanged to another codon encoding the same amino acid

## Value

Creates a character vector of coding nucleotide sequences encoding the same amino acid sequence as the entered sequenceVector. By a mutation rate stated in codonReplacementChance, codons are randomly exchanged, by alternative codons encoding the same amino acid.

## Examples

```
mutatePopulation(c("CGCGATACGCTAAGCGCTACCGATAGTGGA", "TGGGATATTTTAAGCGCTGACGATAGTGGA"), 0.1)
```



---

recombineTwoSequences *Generate new sequence from recombination of two sequences*

---

### Description

This function generates a new sequences through recombination of two parental sequences using 3 modi of recombination. Either random combination of codons, crossover recombination or insertion.

### Usage

```
recombineTwoSequences(ntSequence1, ntSequence2, preferenceVector)
```

### Arguments

ntSequence1      Character value of a nucleotide sequence  
ntSequence2      Character value of a nucleotide sequence  
preferenceVector      Numeric vector of length three which indicates which modus of recombination should be preferred. The first number states the chance of random recombination, the second number indicates the chance of cross-over recombination and the third number indicates the chance of insertion recombination.

### Value

Character value of a nucleotide sequence, generated by recombination from the entered ntSequence1 and ntSequence2. Modes of recombination are cross-over, insertion and random and mode preferences can be stated by preferenceVector.

### Examples

```
recombineTwoSequences("AGGGCCTGGAGGAGGCTT", "TAAGGCAAGCCTGGACCC", c(1, 3, 2))
```

---

selectBestAndMean      *Select best HZEI and mean*

---

### Description

From all sequences of a generation report highest HZEI integral and mean HZEI integral of all.

### Usage

```
selectBestAndMean(sequenceVector, clusterName, increaseHZEI=TRUE)
```

**Arguments**

sequenceVector Character vector of nucleotide sequences  
 clusterName Name of cluster generated with package parallel  
 increaseHZEI Logical value if HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.

**Value**

Numeric vector of length 2 stating the best HZEI integral and the mean HZEI integral of a nucleotide sequence vector sequenceVector. Depending on the increaseHZEI mode, the best HZEI integral value is either the highest (for increaseHZEI==TRUE) or lowest (for increaseHZEI==FALSE).

**Examples**

```
## Setup cluster
library(parallel)
nCores <- 1
clust <- makeCluster(nCores)
clusterExport(clust, list('getOverlappingVectorsFromVector', 'hex',
'calculateHZEIint'), envir = environment())
selectBestAndMean(c('CGCGATACGCTAAGCGCTACCGATAGTGGGA', 'TGGGATATTTAAGCGCTGACGATAGTGGGA'),
clust, increaseHZEI=TRUE)
```

---

selectMatingIndividuals

*Selecting mating sequences from a pool of sequences*

---

**Description**

Selecting sequences from a pool of nucleotide sequences based in chance and their HZEI integral.

**Usage**

```
selectMatingIndividuals(inputGeneration, whoMatesBestPercent=40, whoMatesSemiRandom=20,
whoMatesLuckyly=5, clust, increaseHZEI=TRUE)
```

**Arguments**

inputGeneration  
 Character vector of nucleotide sequences  
 whoMatesBestPercent  
 Numeric value e.g. 20 (which would mean that sequences with the top 20 percent highest HZEI integral are selected for mating)  
 whoMatesSemiRandom  
 Numeric value (is always lower than total number of sequences in input\_generation)  
 whoMatesLuckyly  
 Numeric value (is always lower than total number of sequences in input\_generation)

clust	Name of cluster generated with package parallel
increaseHZEI	Logical value of HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.

**Value**

Character vector of nucleotide sequences which are selected from an entered vector of nucleotide sequences inputGeneration for creation of filial sequences by recombination. Sequences are selected by different criteria stated by whoMatesBestPercent, whoMatesSemiRandom, whoMatesLuckily and increaseHZEI.

**Examples**

```
## Setup cluster
library(parallel)
nCores <- 1
clust <- makeCluster(nCores)
clusterExport(clust, list('getOverlappingVectorsFromVector',
'hex'), envir=environment())
selectMatingIndividuals(c('CGCGATACGCGCGATACG', 'CGCGATACGTGGGATATT',
'CTAAGCGCTCGCGATACG', 'CGCGATACGTTAAGCGCT', 'GACGATAGTCGCGATACG'),
whoMatesBestPercent=40, whoMatesSemiRandom=1, whoMatesLuckily=1, clust, increaseHZEI=TRUE)
```

---

```
slidingWindowHZEImanipulation
```

*Quickly manipulate HZEI integral of nucleotide sequence*

---

**Description**

Quickly manipulate HZEI integral of nucleotide sequence (min. 21nt long)

**Usage**

```
slidingWindowHZEImanipulation(inSeq, increaseHZEI=TRUE)
```

**Arguments**

inSeq	Character value of nucleotide sequence (min 21nt long, only bases 'A', 'G', 'T' or 'C')
increaseHZEI	Logical value if HZEI integral should be increased or decreased during SD degradation. If TRUE, function aims to increase HZEI integral.

**Value**

Character value of a nucleotide sequence encoding the same amino acid sequence as inSeq, but an increased HZEI integral, due to alternative codon selection, accomplished through sliding window optimization.

**Examples**

```
# Load R packages
library('parallel')
library('utils')
library('data.table')

# Set parameters for genetic algorithm
inSeq <- 'ATGGAAGACGCCAAAAACATAAAGAAAGGCAGGCTAAGCCTAGCTTGCCATTGCCCGGCGCCATTCTATCCGCTGGAAGATGGAATT'

maximizedHZEIseq <- slidingWindowHZEImanipulation(inSeq, increaseHZEI=TRUE)
minimizedHZEIseq <- slidingWindowHZEImanipulation(inSeq, increaseHZEI=FALSE)

#Access sequence with maximized HZEI integral
maximizedHZEIseq

#Access sequence with minimized HZEI integral
minimizedHZEIseq
```

---

startModConApp	<i>Start GUI of VarCon.</i>
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---

**Description**

Start graphical user interface for the ModCon application.

**Usage**

```
startModConApp()
```

**Value**

Shiny app

**Examples**

```
startModConApp()
```

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