# Estimating cell type composition in whole blood using <br> differentially methylated regions 

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Bioconductor 2017

## What is DNA Methylation?

m m
m


## TAGCGCAATGTCGCCTT <br> | |

$u m$
u

## What is DNA Methylation?

m m
AT ${ }^{\bullet} G{ }^{\bullet}{ }^{\text {CGTTACTGCGGAA }}$
TAGCGCAATGTCGCCTT

I
u

$\downarrow$

## What is DNA Methylation?

m
ATCGCGTTACTGGGGAA

$$
\begin{gathered}
\text { TAGCGCAATGTCGCCTT } \\
\mid \\
\mathrm{m} \\
\mid \\
\hline \mathrm{l}
\end{gathered}
$$

## DNA methylation in whole blood correlates with age at this one CpG



Data from GSE32148

## Blood is a mixture of many cell types

## Whole blood cell types:

- Tcells
- CD8T
- CD4T
- Natural Killer
- Bcells
- Granulocytes
- Monocytes


## Bioconductor data package available:

- Data originally from Reinius et al. (2012)
> library(FlowSorted.Blood.450k)



## Cell composition changes with age



## Statistical Model: Houseman et al. (2012)


$i=(1, \ldots, N)=$ whole blood samples
$j=(1, \ldots, J)=$ CpGs
$k=(1, \ldots, K)=$ cell type profiles
whole blood sample

relative cell type proportions

Measurement error


## New platform technologies emerging

## First approach

- Apply Houseman method using new platform technology

Problems with this approach

1. Observed methylation levels depend on platform used
2. Not all CpGs are included in new platforms

## Platform-dependent differences between 450k array and RRBS platforms

Chromosome 6


## Platform-dependent differences between 450k array and RRBS platforms



## New platform technologies emerging

## First approach

- Apply Houseman method using new platform technology


## Problems with this approach

1. Observed methylation levels depend on platform
2. Not all CpGs are included in new platforms

## Cell types preserve their methylation state across regions

Chromosome 14
Beta values
(Purified cell types on measured on 450k array)

- Identify regions using bumphunter BioC pkg



## Recall Houseman Model:

$$
Y_{i j}=\sum_{k=1}^{K} \pi_{i k} X_{j k}+\varepsilon_{i j} \quad \begin{aligned}
& i=(1, \ldots, N)=\text { whole blood samples } \\
& j=(1, \ldots, J)=\text { CpGs } \\
& k=(1, \ldots, K)=\text { cell type profiles }
\end{aligned}
$$



## Our proposed model:

$Y_{r}=\sum_{k=1}^{K} \pi_{k}\left[\left(1-Z_{r k}\right) \delta_{o, r}+Z_{r k} \delta_{1, r}\right]+\varepsilon_{r}$

$$
\begin{gathered}
\delta_{0, r} \sim N\left(\alpha_{0}, \sigma_{0}^{2}\right) \\
\delta_{1, r} \sim N\left(\alpha_{1}, \sigma_{1}^{2}\right) \\
\varepsilon_{r} \sim N\left(0, \sigma^{2}\right)
\end{gathered}
$$

$Z_{r k}=\left\{\begin{array}{ccl}1 & \text { if region } r \text { and cell type } k \text { is methylated } & r=(1, \ldots ., R)=\text { differentially methylated regions } \\ 0 & \text { otherwise } & k=(1, \ldots, K)=\text { cell types }\end{array}\right.$


## How does our model perform?

## $N=800$ whole blood samples run on 450 k microarray platform



RMSE:
0.0385

RMSE:
0.0531

Cell composition estimates from whole blood samples measured on two platforms
$N=12$ samples measured on two platforms:

- 450k microarray
- RRBS sequencing



### 3.1 Demo for BioC 2017

## 1ibrary (methylcc)

library(minfi)
library(FlowSorted.Blood.450k)
data(FlowSorted.Blood.450k)
Comparing cell composition estimates

```
# Subset RGChan
rgset <- FlowSo
```

\# Use methy7cC:
est.methylcc <- counts.methy1CC
\# Compare to mi
sampleNames (rgs
counts.minfi <-


## For more information

## methyICC:

https://github.com/stephaniehicks/methylCC

## Comments/Suggestions:

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